

# Taq-Plus PCR Master Mix (2×)

REF: EG15117M

### **Storage Condition**

-20°C

### Components

| Component                    | Amount |
|------------------------------|--------|
| Taq-Plus PCR Master Mix (2×) | 5×1 ml |

## **Description**

Taq-Plus PCR Master Mix  $(2\times)$  is a pre-mixed PCR reaction solution formulated with Taq DNA Polymerase, dNTPs, and all of the components required for PCR, except DNA template and primers. This pre-mixed formulation saves time and reduces contamination due to the fewer pipetting steps required for PCR set up.

It is capable of robust amplification of up to 6 kb from genomic DNA. PCR products amplified with this mix have a 3'-terminal adenosine (A), and therefore PCR products can be used directly for T/A cloning.

### **Quality Control Assays**

#### **Endonuclease Activity**

A 50  $\mu$ I reaction containing 200 ng of supercoiled plasmid and 25  $\mu$ I of Taq-Plus PCR Master Mix incubated for 4 hours at 37°C results in <10% conversion to the nicked or linearized form as determined by agarose gel electrophoresis.

#### **Non-specific Nuclease Activity**

A 50  $\mu$ I reaction containing 15 ng of dsDNA fragments and 25  $\mu$ I of Taq-Plus PCR Master Mix incubated for 16 hours at 37 °C results in no detectable degradation of the dsDNA fragments as determined by agarose gel electrophoresis.

#### **Protocol**

#### 1. PCR Reaction System

| Reagent                             | Amount   | Final Concentration |
|-------------------------------------|----------|---------------------|
| Taq-Plus PCR Master Mix (2×)        | 25 µl    | 1×                  |
| Forward Primer (10 µM) <sup>a</sup> | 1~2 µl   | 0.2~0.4 μM          |
| Reverse Primer (10 µM) <sup>a</sup> | 1~2 µl   | 0.2~0.4 μM          |
| Template DNA <sup>b</sup>           | xμl      |                     |
| ddH₂O                               | To 50 μl |                     |

- a. Recommended final concentration for primers is 0.2  $\mu M.$  Adjustments can be made in the range of 0.1~1  $\mu M.$
- b. Optimal reaction concentrations may vary for different templates. For a 50  $\mu$ l reaction system, the recommended template amounts are approximately 10~400 ng for genomic DNA and 10 pg~20 ng for plasmid or viral DNA.

#### 2. PCR Conditions

| Step                              | Temperature     | Time       |              |
|-----------------------------------|-----------------|------------|--------------|
| Initial denaturation <sup>a</sup> | 94°C            | 3~5 min    |              |
| Denaturation                      | 94°C            | 30 s       | •            |
| Annealing                         | <b>55~65</b> °C | 30 s       | 30~35 Cycles |
| Extension <sup>b</sup>            | <b>72</b> °C    | 30~60 s/kb |              |
| Final Extension                   | <b>72</b> °C    | 5 min      |              |

- a. The initial denaturation condition is suitable for most amplification reactions. For some complex templates, such as bacteria and yeast colon, the initial denaturation time can be extended to 10 minutes to improve denaturation efficiency.
- b. It is recommended to set at 30 s/kb when the target fragment is shorter than 2 kb. If the length of the target fragment exceeds 2 kb, it is recommended to set at 60 s/kb.

Note: If yeast cell is used as the PCR template, it is recommended that the target fragment length does not exceed 2.5 kb. If it exceeds 2.5 kb, it is advisable to use the lysed cells as template.