

BbvCI

REF: EG25516S



Storage Condition

-20°C

Components

Component	Amount
BbvCI (2 U/μl)	25 μl
10× CutOne® Buffer	1 ml
10× CutOne® Color Buffer	1 ml

Description

BbvCI is derived from the BbvCI gene of *Bacillus brevis* (L. Ge) and is obtained through recombinant expression in *Escherichia coli*. BbvCI is a Type IIA restriction enzyme existing as a heterodimer composed of two distinct catalytic subunits, each with an independent cleavage site. It recognizes and cleaves the non-palindromic sequence CCTCAGC(-5/-2), generating 5' cohesive ends with 3-base overhangs. BbvCI uses CutOne® reaction buffer and is compatible with double digestion with other LightNing® restriction enzymes. However, it is advisable to avoid extended digestion (exceeding 3 hours) to prevent star activity.

BbvCI shows superior activity in the universal CutOne® and CutOne® Color Buffer. CutOne® Color Buffer includes a density reagent along with red and yellow tracking dyes that allow for direct loading of the reaction mixtures on a gel. The red dye of the CutOne® Color Buffer migrates with 2.5 kb double-strand DNA fragments in a 1% agarose gel, and the yellow dye migrates with 10 bp double-strand DNA fragments in a 1% agarose gel.

Recommended Reaction Conditions

1× CutOne® Buffer;

Incubate at 37°C ;

Refer to "Protocol for DNA Digestion" for reaction setup.

Heat Inactivation

Incubation at 80°C for 20 minutes.

Definition of Activity Unit

One unit of activity refers to the amount of enzyme required to completely digest 1 μg of λDNA in a 50 μl reaction system at 37°C for 1 hour.

Quality Control Assays

Function

2 U of BbvCI can completely digest 1 μg of λDNA within 15 minutes at 37°C .

Prolonged Incubation / Star Activity Assay

Under optimal reaction temperature, incubate 2 U BbvCI with 1 μg λDNA for 3 hours. No contamination from other nucleases or non-specific substrate degradation caused by star activity was detected. Longer incubation may result in star activity.

Ligation and Recutting

Under optimal reaction temperature, digest the substrate using 2 U BsrDI and recover the digested products. <10% of the DNA fragments can be ligated with T4 DNA Ligase at 22°C . Of these ligated fragments, >95% can be recut with BsrDI as determined by agarose gel electrophoresis.

Icon Descriptions

- The enzyme's optimum reaction temperature is 37°C .
- Cleavage with this restriction enzyme may be blocked or impaired when the substrate DNA is methylated by the CpG methylase.
- The enzyme can be heat inactivated at by incubation 80°C for 20 minutes.
- 3 hours incubation did not show star activity, but delayed enzyme digestion might show star activity.

Protocol

1. Protocol for DNA Digestion

① Combine the following components on ice in the following order:

ddH ₂ O	up to 50 μl
10× CutOne® Buffer or 10× CutOne® Color Buffer	5 μl
DNA ^a	1 μg
BbvCI (2 U/μl)	1 μl
Total	50 μl

a. DNA substrates should contain no phenol, chloroform, ethanol, EDTA, detergents, or high salt concentrations, otherwise enzyme activity will be affected;

- ② Mix gently and spin down.
- ③ Incubate at 37°C for 1~3 hours.
- ④ Optional: Inactivate the enzyme by heating at 80°C for 20 minutes, or by adsorption column or phenol/chloroform purification to terminate the reaction.

2. Notice

- ① The volume of enzyme added to the reaction mixture should not exceed 10% of the total volume to avoid star activity caused by excessive glycerol in the enzyme storage buffer.
- ② The additives (e.g., glycerol, salt) in the enzyme storage buffer are the same as the contaminants in the substrate solution (e.g., salt, EDTA, or ethanol, etc.). Therefore, the smaller the reaction volume, the stronger the digestion inhibition effect.

Number of Recognition Sites in DNA

λDNA	ΦX174	pBR322	pUC57	pUC18/19	SV40	M13mp18/19	Adeno2
7	3	0	0	0	0	2	9

Methylation Effects on Digestion

Dam	Dcm	CpG	EcoKI	EcoBI
No effect	No effect	Impaired	No effect	No effect

Activity in Different Buffers*

	CutOne® Buffer	Thermo Scientific FastDigest Buffer	NEB rCutSmart™ Buffer	Takara QuickCut™ Buffer
Activity	100%	100%	100%	100%

*The activity data come from the functional test described above.